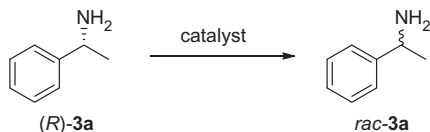


Figure 1. (a) HR STEM image of Pd on AlO(OH) (**2**). (b) Size distribution of Pd NPs obtained by HR STEM analysis.

Table 1
Racemization of (*R*)-1-methylbenzylamine (**3a**)^a



Entry	Catalyst	Amount (mol %)	Temp (°C)	Time (h)	ee ^b (%)
1 ^c	Ru complex	8	90	24	55
2 ^d	Pd/BaSO ₄	5.8	70	24	2
3 ^e	1	1	70	24	2
4	2	1	70	3	4
5	2	1	50	12	7
6	2	1	40	24	8

^a Conditions: 0.1 mmol (*R*)-**3a**, 1 mL of toluene.

^b Determined by chiral HPLC.

^c Data from Ref. 7b. The Ru complex employed is dimeric (4 mol %) but the active species is monomeric (8 mol %).

^d Data from Ref. 4a.

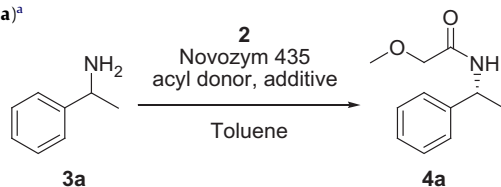
^e Data from Ref. 5a.

no rationale is available for such a high activity of **2**. We however speculate that the smaller size and larger surface area of Pd NPs would be a major contributing factor. Also the method of preparation would affect the activity of Pd. Catalyst **2** was prepared by

impregnating the dried AlO(OH) matrix with Pd NPs while **1** was prepared by entrapping Pd NPs into the AlO(OH) gel during gelation. Accordingly, the substrates would be more readily accessible to Pd NPs of the former than those of the latter.

The performance of **2** as the racemization catalyst in DKR was explored with a commercial lipase (Novozym 435) as the resolution catalyst. The DKR reactions of racemic **3a** (0.3 mmol) were performed under several different conditions for optimization (Table 2).¹⁴ In the presence of **2** (1 mol % of Pd), Novozym 435 (25 mg/mmol of substrate), and ethyl methoxyacetate (1.7 equiv) as the acyl donor in toluene at 70 °C, the DKR took 24 h for completion but the yield (85%) was less than satisfactory (entry 1). The use of isopropyl methoxyacetate as the acyl donor improved the yield to 90% (entry 2). It is believed that isopropanol from isopropyl methoxyacetate during DKR facilitates the racemization of amines by acting as an additional hydrogen source for the returning of imines to amines.^{4a,7b} A further improvement in reaction time from 24 to 12 h was achieved with the use of sodium carbonate as a base to trap any acidic species from the commercial enzyme preparation or the acyl donor hydrolyzed (entry 3). Previously, the corresponding DKR with **1** took 72 h to give similar yield and ee.^{5a} Therefore, the present DKR is sixfold faster compared to the previous DKR. Finally, we were able to further reduce the reaction time to 6 h by doubling the amounts of catalysts (entry 4). The optimized DKR afforded a high yield (92%) and an excellent enantiopurity

Table 2
Dynamic kinetic resolution of 1-methylbenzylamine (**3a**)^a



Entry	Pd (mol %)	Lipase (mg/mmol)	Na ₂ CO ₃ (mg/mmol)	Temp (°C)	Time (h)	Conv ^b (%)	Yield ^c (%)	ee ^d (%)
1 ^e	1	25	0	70	24	97	85	99
2 ^f	1	25	0	70	24	96	90	99
3 ^f	1	25	25	70	12	97	85	99
4 ^f	2	50	50	70	6	>97	92	99
5 ^f	1	25	25	50	48	>97	90	98
6 ^f	2	25	25	40	48	>97	98	98

^a Conditions: 0.3 mmol **3a**, 1.7 equiv of acyl donor, 6 mL of toluene.

^b Determined by ¹H NMR.

^c Isolated yield.

^d Determined by chiral HPLC.

^e Acyl donor = ethyl methoxyacetate.

^f Acyl donor = isopropyl methoxyacetate.

Table 3
Dynamic kinetic resolution of additional benzyl amines^a

Entry	Substrate	Pd (mol %)	Novozym (mg/mmol)	Time (h)	yield ^b (%)	ee ^c %
1		5	50	6	94	98
2 ^d		4	50	6	99	98
3		5	50	6	88	97
4		5	50	6	97	>99
5		5	50	6	96	>99
6		5	50	6	83	90
7		5	40	12	93	96
8		2	150	6	89	>99
9		2	150	12	97	>99

^a Conditions: 0.3 mmol of substrate, 1.7 equiv of isopropyl methoxyacetate, 6 mL of toluene, 70 °C, 6 h.

^b Isolated yield.

^c Determined by chiral HPLC.

^d Using methyl methoxyacetate as an acyl donor.

(99% ee). The DKR reactions at lower temperatures (40–50 °C) also proceeded smoothly within a reasonable reaction time (48 h) to provide excellent results (entries 5 and 6). This is important because the amine DKR would become possible with other enzymes, which lose their activities at 70 °C or higher but have acceptable stability at 40–50 °C.

To see the scope and generality of the DKR procedure using **2** as the racemization catalyst, we carried out the DKR reactions of additional benzyl amines **3b–h** (Table 3). It was found that ring-substituted benzyl amines **3b–g** were less prone to racemization than **3a**¹⁵ so that they needed 4–5 mol % of **2** for the fast DKR within 6 h. Interestingly, **3b** with an electron-withdrawing substituent on benzene ring underwent its DKR with high yield (99%) and excellent ee (98%), whereas the DKR of **3d** with a strongly electron-donating substituent provided a lower yield (88%) with some byproducts^{4a} resulting from the reactions of amine substrates with their racemization intermediates (imines)¹⁶ (entries 2 and 3).

The DKRs of two six-membered alicyclic amines (**3e** and **3f**) proceeded smoothly well to afford high yields and excellent enantiopurities (entries 4 and 5). The DKR of five-membered alicyclic amine **3g**, however, was less than satisfactory in both yield and enantiopurity (entry 6). To obtain better results, the reaction time was increased to 12 h with a decrease in the amount of enzyme (entry 7). The DKR of a slow-reacting substrate (**3h**) did not proceed to completion in 6 h even though a three times larger amount of enzyme was employed (entry 8). The reaction time thus was

extended to 12 h to obtain a high yield with an excellent enantiopurity (entry 9).

In summary, we have demonstrated that the fast racemization and DKR of 1-methylbenzylamine can be achieved with a Pd nanocatalyst containing Pd nanoparticles smaller than 2 nm in average diameter, and the DKR procedure is applicable to other benzyl amines. It is noteworthy that the DKR can be carried out at a mild temperature (40 °C) which is significantly lower than the previously reported cases. Thus it should be possible to use other enzymes, which are less thermostable than Novozym 435, with the Pd nanocatalyst for the DKR of amines. Further studies toward this end are under way at this laboratory.

Acknowledgments

This work was supported by the National Research Foundation of Korea (2009–0087801) and POSCO. The authors thank the Korean Government for supporting our graduate program via the BK21 project.

References and notes

- Reviews: (a) Kim, M.-J.; Ahn, Y.; Park, J. *Curr. Opin. Biotechnol.* **2002**, *13*, 578–587; (b) Pamies, O.; Bäckvall, J. E. *Chem. Rev.* **2003**, *103*, 3247–3262; (c) Kim, M.-J.; Ahn, Y.; Park, J. *Bull. Korean Chem. Soc.* **2005**, *26*, 515–522; (d) Martin-Matute, B.; Bäckvall, J. E. *Curr. Opin. Biol.* **2007**, *11*, 226–232; (e) Ahn, Y.;

- Ko, S.-B.; Kim, M.-J.; Park, J. *Coord. Chem. Rev.* **2008**, *252*, 647–658; (f) Lee, J. H.; Han, K.; Kim, M.-J.; Park, J. *Eur. J. Org. Chem.* **2010**, 999–1015.
2. (a) Choi, J. H.; Kim, Y. H.; Nam, S. H.; Shin, S. T.; Kim, M.-J.; Park, J. *Angew. Chem., Int. Ed.* **2002**, *41*, 2373–2376; (b) Choi, J. H.; Choi, Y. K.; Kim, Y. H.; Park, E. S.; Kim, E. J.; Kim, M.-J.; Park, J. *J. Org. Chem.* **2004**, *69*, 1972–1977; (c) Martin-Matute, B.; Edin, M.; Bogar, K.; Bäckvall, J. E. *Angew. Chem., Int. Ed.* **2004**, *43*, 6535–6539; (d) Kim, N.; Ko, S.-B.; Kwon, M. S.; Kim, M.-J.; Park, J. *Org. Lett.* **2005**, *7*, 4523–4526; (e) Berkessel, A.; Sebastian-Ibarz, M. L.; Müller, T. N. *Angew. Chem., Int. Ed.* **2006**, *45*, 6567–6570; (f) Akai, S.; Tanimoto, K.; Kanao, Y.; Egi, M.; Yamamoto, T.; Kita, Y. *Angew. Chem., Int. Ed.* **2006**, *45*, 2592–2595; (g) Haak, R. M.; Berthiol, F.; Jerphagnon, T.; Gayet, J. A.; Tababionio, C.; Postema, C. P.; Ritleng, V.; Pfeffer, M.; Janssen, D. B.; Minnaard, A. J.; Feringa, B. L.; De Vries, J. G. *J. Am. Chem. Soc.* **2008**, *130*, 13508–13509; (h) Kim, M.-J.; Choi, Y. K.; Kim, S.; Kim, D.; Han, K.; Ko, S.-B.; Park, J. *Org. Lett.* **2008**, *10*, 1295–1298.
3. (a) Reetz, M. T.; Schimossek, K. *Chimia* **1996**, *50*, 668–669; (b) Choi, Y. K.; Kim, M. J.; Ahn, Y.; Kim, M.-J. *Org. Lett.* **2001**, *3*, 4099–4101.
4. (a) Parvulescu, A.; De Vos, D.; Jacobs, P. *Chem. Commun.* **2005**, 5307–5309; (b) Parvulescu, A. N.; Jacobs, P. A.; De Vos, D. E. *Chem. Eur. J.* **2007**, *13*, 2034–2043; (c) Andrade, L. H.; Silva, A. V.; Pedrozo, E. C. *Tetrahedron Lett.* **2009**, *50*, 4331–4334.
5. (a) Kim, M.-J.; Kim, W.-H.; Han, K.; Choi, Y. K.; Park, J. *Org. Lett.* **2007**, *9*, 1157–1159; (b) Han, K.; Park, J.; Kim, M.-J. *J. Org. Chem.* **2008**, *73*, 4302–4304; (c) Choi, Y. K.; Kim, Y.; Han, K.; Park, J.; Kim, M.-J. *J. Org. Chem.* **2009**, *74*, 9543–9545.
6. Parvulescu, A. N.; Jacobs, P. A.; De Vos, D. E. *Adv. Synth. Catal.* **2008**, *350*, 113–121.
7. (a) Pamies, O.; Ell, A. H.; Samec, J. S. M.; Hermans, N.; Bäckvall, J. E. *Tetrahedron Lett.* **2002**, *43*, 4699–4702; (b) Paetzold, J.; Bäckvall, J. E. *J. Am. Chem. Soc.* **2005**, *127*, 17620–17621; (c) Veld, M. A. J.; Hult, K.; Palmans, A. R. A.; Meijer, E. W. *Eur. J. Org. Chem.* **2007**, 5416–5421; (d) Thalén, L. K.; Zhao, D.; Sortais, J.-B.; Paetzold, J.; Hoben, C.; Bäckvall, J. E. *Chem. Eur. J.* **2009**, *15*, 3403–3410.
8. (a) Stirling, M.; Blacker, J.; Page, M. I. *Tetrahedron Lett.* **2007**, *48*, 1247–1250; (b) Blacker, A. J.; Stirling, M. J.; Page, M. I. *Org. Process Res. Dev.* **2007**, *11*, 642–648.
9. Recently, a faster amine DKR using a sulfanyl radical for racemization was reported. (a) Gastaldi, S.; Escoubet, S.; Vanthuynne, N.; Gil, G.; Bertrand, M. P. *Org. Lett.* **2007**, *9*, 837–839; (b) Routaboult, L.; Vanthuynne, N.; Gastaldi, S.; Gil, G.; Bertrand, M. P. *J. Org. Chem.* **2008**, *73*, 364–368; (c) Blidi, L. E.; Nechab, M.; Vanthuynne, N.; Gastaldi, S.; Bertrand, M. P.; Gil, G. *J. Org. Chem.* **2009**, *74*, 2901–2903.
10. Heiz, U.; Landman, U. *Nanocatalysis*; Springer: Berlin, Heidelberg, New York, 2006.
11. (a) Tsunoyama, H.; Sakurai, H.; Negishi, Y.; Tsukuda, T. *J. Am. Chem. Soc.* **2005**, *127*, 9374–9375; (b) Evans, G.; Kozhevnikov, I. V.; Kozhevnikova, E. F.; Claridge, J. B.; Vaidyanathan, R.; Dickinson, C.; Wood, C. D.; Cooper, A. I.; Rosseinsky, M. J. *J. Mater. Chem.* **2008**, *18*, 5518–5523; (c) Beck, I. E.; Bukhtiyarov, V. I.; Pakharukov, I. Y.; Zaikovskiy, V. I.; Kriventsov, V. V.; Parmon, V. N. *J. Catal.* **2009**, *268*, 60–67; (d) Wang, C.; van der Vliet, D.; Chang, K.-C.; You, H.; Strmcnik, D.; Schlueter, J. A.; Markovic, N. M.; Stamenkovic, V. R. *J. Phys. Chem. C* **2009**, *113*, 19365–19368.
12. *Preparation of AIO(OH)*. (sec-BuO)₂Al (5.0 mL, 19.5 mmol) and 2-butanol (10 mL) were added to a 50 mL round-bottomed flask and then stirred at room temperature for 10 min to obtain a homogeneous solution, followed by the addition of water (3 mL). The reaction mixture was stirred at room temperature for 20 min followed by the addition of acetone (30 mL) and then stirred for additional 30 min. The mixture was filtered, washed with acetone, and dried at 150 °C under a reduced pressure to give AIO(OH) as white powder (1.54 g). *Preparation of 2*. Pd(OAc)₂ (33 mg, 0.147 mmol) and ethanol (10 mL, 170 mmol) were added to a 50 mL round-bottomed flask and stirred at room temperature for 10 min followed by the addition of AIO(OH) (1.54 g). The resultant mixture was further stirred at room temperature for 1 h to allow the spontaneous reduction of Pd(II) to Pd(0) nanoparticles adsorbed on AIO(OH) powder. The heterogeneous mixture was filtered, washed with acetone, and dried at 80 °C under a reduced pressure to give **2** as dark powder (Pd on AIO(OH); 1.46 g). The ICP analysis indicated that the Pd nanocatalyst contained 0.85 wt % of Pd.
13. *Racemization of (R)-3a*. (R)-**3a** (12 mg, 0.10 mmol) was added to a suspension of Pd catalyst **2** (13 mg, 1.0 mol % of Pd) in dry and degassed toluene (1 mL; 0.10 M). The reaction mixture was stirred at 70 °C under argon. After 3 h, the reaction mixture was cooled to room temperature and filtered through a glass filter (pore size: 20–30 μm). The filtrate was concentrated and analyzed by ¹H NMR spectroscopy and HPLC (Kromasil-5-CellCoat, *n*-hexane/2-propanol/diethyl amine = 90:10:0.1, flow rate = 1.5 mL/min, UV 220 nm): (R)-**3a** = 5.57 min, (S)-**3a** = 6.57 min.
14. *Dynamic kinetic resolution of 3a*. A solution containing **3a** (36 mg, 0.30 mmol), Pd nanocatalyst **2** (76 mg, 2.0 mol % of Pd), Novozym 435 (15 mg), sodium carbonate (15 mg), and isopropyl methoxyacetate (71 μL, 1.7 equiv) in dry and degassed toluene (6 mL) was stirred at 70 °C under argon atmosphere. After 6 h, the reaction mixture was cooled to room temperature and filtered through a glass filter with celite pad. The filtrate was concentrated and analyzed by ¹H NMR spectroscopy, indicating that all of the substrate was consumed. The mixture was subjected to a flash column chromatography (silica gel, *n*-hexane/ethyl acetate = 1:1) to provide **4a** (53.6 mg, 0.277 mmol, 92%, 99% ee): mp 61–63 °C (lit.^{5a} mp 59–60 °C); [α]_D²⁵ + 90.1 (c 1.0, CHCl₃) (lit.^{5a} [α]_D²⁵ + 85.2 (c 0.5, CHCl₃)); HPLC ((R,R)Whelk-O1, *n*-hexane/2-propanol = 80:20, flow rate = 2.0 mL/min, UV 217 nm): (S)-**4a** = 7.06 min, (R)-**4a** = 17.08 min; ¹H and ¹³C NMR data are in good agreement with the literature data.^{5a}
15. In general, the racemization of benzyl amines takes place through a two-step process, dehydrogenation and hydrogenation via an imine intermediate. The observation that the racemizations of **3b–g** took place slowly relative to that of **3a** in DKR indicates that racemization is retarded by both electron-donating and electron-attracting groups on the benzene ring.
16. In racemization, the dehydrogenation step should be rate determining with electron-attracting groups while the hydrogenation step should be rate determining with electron-donating groups. Accordingly, the life time of imine intermediate in the latter case is long relative to that in the former case. This explains the reason why the amounts of byproducts resulting from the side reactions of imine intermediate (including its reaction with amine substrate) are significantly larger in the DKR of **3d** with an electron-donating group than in the DKR of **3b** with an electron-attracting group.